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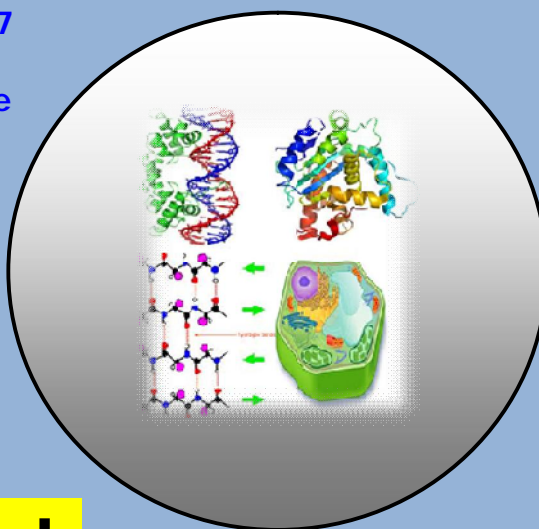
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RESEARCH PAPER

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Antifungal Activity of Some Latex against Seed-Borne Pathogenic Fungi

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ABSTRACT

Plant extracts and plant products are being used to control the diseases since last several years. Latex of the various plants is found to be effective against seed-borne pathogenic fungi. The in vitro studies have been performed by using cup-plate method to examine the antifungal activity of some latex. Latex of 9 plants was screened against 5 seed-borne pathogenic fungi viz. Alternaria alternata, Aspergillus niger, Curvularia lunata, Fusarium moniliforme and Trichoderma viride. Latex of 5 plants showed antifungal activity. The latex of Jatropha curcas showed maximum activity (Mean activity zone 18.20 mm); while minimum activity was observed with latex of Carica papaya (Mean activity zone 14.60 mm). The latex collected from Alstonia scholaris, Ficus benghalensis, F. elastic and F. racemosa was unable to exhibit any activity against the fungi under investigation. These plant extracts can possibly be exploited in the management of seed-borne pathogenic fungi to prevent biodeterioration of seeds in an eco-friendly way.

Key Words: *Antifungal activity, Seed-borne Pathogenic Fungi, Biodeterioration, and Latex.*

INTRODUCTION

Fungal diseases are known to cause great damages all over the world. Different species of *Alternaria*, *Aspergillus*, *Ceratobasidium*, *Cercospora*, *Cochliobolus*, *Curvularia*, *Drechslera*, *Fusarium*, *Gaeumannomyces*, *Microdochium*, *Penicillium*, *Pyricularia*, *Pythium*, *Rhizoctonia*, *Rhizopus*, *Sclerophthora*, *Trichoderma* and *Tricoconella* are most common associates of seeds all over the world, causing pre and post-infections and considerable quality losses viz. seed abortion, seed rot, seed necrosis, reduction or elimination of germination capacity, seedling damage and their nutritive value have been reported (Miller, 1995; Janardhana *et al.*, 1998; Kavitha *et al.*, 2005). Seed treatment is the safest and the cheapest way of control of seed-borne fungal diseases and to prevent biodeterioration of grains (Chandler, 2005; Bagga and Sharma, 2006).

Even though effective and efficient control of seed-borne fungi can be achieved by the use of synthetic chemical fungicides, the same cannot be applied to grains for reasons of pesticide toxicity (Harris *et al.*, 2001). The toxic effect of synthetic chemicals can be overcome, only by persistent search for new and safer pesticides accompanied by wide use of pest control methods, which are eco-friendly and effective (Mohana *et al.*, 2011). Green plants represent a reservoir of effective chemotherapeutants and can provide valuable sources of natural pesticides (Balandrin *et al.*, 1985; Hostettmann and Wolfender, 1997). The latex is widely used in cosmetics, pharmaceuticals, food, paper, textile and petroleum industries (Madhavi *et al.*, 2013). Daruliza *et al.*, (2011) reported that the *Hevea brasiliensis* latex C-serum exerted a specific antifungal activity against *Aspergillus niger*. Hussain *et al.*, (2014) performed an *in-vitro* comparative study for antimicrobial activity of fresh latex, juice and extract of *Euphorbia hirta* and *Euphorbia thymifolia*. They concluded that the fresh latex of the *E. hirta* is a potentially good candidate for the therapy of antibacterial resistant bacteria and suggested further study. Saadabi *et al.*, (2012) evaluated leaves, stems, fruits and latex of *Calotropis procera* extracts against four types of bacteria and two types of fungi i.e. *Aspergillus niger* and *Candida albicans*. They clearly noticed that the latex of the plant had a broad spectrum activity against bacteria and fungi under investigation. Vashist & Jindal (2012) reported antifungal activity of latex of *Calotropis gigantea* against fungi viz. *C. albicans*, *Saccharomyces cerevisiae*, *Trichophyton mentagrophytes*, *Trichophyton rubrum*, *A. fumigatus*, *A. flavus*, *A. niger*, *Penicillium chrysogenum*.

MATERIALS AND METHOD

Fungal pathogens were isolated on PDA medium from different stored seeds. Identified fungal cultures were isolated and pure cultures of each fungi made separately on PDA slants. These pure cultures were used for further investigation.

a) Collection of Latex: Fresh latex of the plants was collected from the fields and used for the study (Pawar, 1999).

b) Cup Plate Method: 20 ml of PDA media was poured in sterilized petridishes (9 cm diameter) and allowed to solidify. Then pure cultures of fungi were streaked out in regular intervals on the media poured in petridishes. In the centre of the medium, a cup cavity of 8 mm diameter was made with sterilized No. 4 cork borer. This cup was filled with 0.1 ml of the stem extract (Pawar and Papdiwal, 2010). The petridishes were incubated for 6 days at $30\pm 2^{\circ}\text{C}$ temperature and the observations were recorded as diameter of inhibitory zone in mm. Cup plate filled with sterile distilled water was used as control in all the experiments (Pawar, 2013). All the experiments were in triplicate and mean has been considered in observation table.

RESULTS AND DISCUSSION

The antifungal activity of latex of 9 plants against 5 seed-borne fungi is presented in table 1 as zone of inhibition (in mm). It was observed from table 1 that out of 9 latex tested, latex of 5 plants showed antifungal activity; out of which *Jatropha curcas* showed maximum activity (Mean activity zone 18.60 mm), followed by latex of *Calotropis gigantea* (Mean activity zone 18.20 mm) and minimum activity was observed with *Carica papaya* latex (Mean activity zone 14.60 mm).

While latex of *Michelia champaca* and *Moringa oleifera* also showed good antifungal activity; however, latex of *Alstonia scholaris*, *Ficus benghalensis*, *Ficus elastic* and *Ficus recemosa* could not show any antifungal activity against the fungi under investigation.

Table 1. Antifungal activity of Root Extracts against Seed-borne Pathogenic Fungi.

Sr. No	Name of the Plant	Zone of Inhibition (in mm)					
		<i>Alternaria alternate</i>	<i>Aspergillus niger</i>	<i>Curvularia lunata</i>	<i>Fusarium moniliforme</i>	<i>Trichoderma viride</i>	Mean
1	<i>Alstonia scholaris</i> (L.) R. Br.	–	–	–	–	–	–
2	<i>Calotropis gigantea</i> (L.) R. Br.	18	19	17	18	19	18.20
3	<i>Carica papaya</i> L.	15	14	14	14	16	14.60
4	<i>Michelia champaca</i> L.	15	16	15	15	16	15.40
5	<i>Ficus benghalensis</i> L.	–	–	–	–	–	–
6	<i>Ficus elastica</i> Roxb.	–	–	–	–	–	–
7	<i>Ficus recemosa</i> L.	–	–	–	–	–	–
8	<i>Jatropha curcas</i> L.	18	19	18	18	20	18.60
9	<i>Moringa oleifera</i> Lamk.	17	16	17	18	16	16.80

–: No Activity.

Similar results were reported by Kumar *et al*, (2010). They tested antimicrobial activity of crude aqueous extract of latex of *Calotropis gigantean* against six pathogenic species of bacteria and two species of fungi viz. *C. krusei* and *A. niger*. Murugan (2012) also reported antimicrobial activity of leaves extract and latex of *C. gigantean*. Sharma *et al*, (2010) reported broad-spectrum antimicrobial properties of *Jatropha curcas*. Mbakwem-Aniebo *et al*, (2012) also studied effects of *J. curcas* leaves on common Dermatophytes and causative agent of Pityriasis versicolor in Rivers State of Nigeria

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